

Proximate compositions, bioactive compounds and antioxidant activity from large-leafed mangrove (*Bruguiera gymnorrhiza*) fruit

^{1*}Sudirman, S., ²Nurjanah and ²Jacoeb, A. M.

¹Fisheries Product Technology, Faculty of Agriculture, Sriwijaya University, Indralaya, Ogan Ilir (OI) – 30669 Palembang, South Sumatra, Indonesia

²Department of Aquatic Product Technology, Faculty of Fisheries and Marine Sciences, Bogor Agricultural

University

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Keywords

Antioxidant Bioactive compound Large-leafed mangrove Chromatography Antioxidants are compounds that can inhibit or prevent the oxidation of the easily oxidized substrate. One of the plants as a potential source of bioactive compounds and antioxidant activity is large-leafed mangrove (*Bruguiera gymnorrhiza*). This plant was commonly found in the Pacific region of Southeast Asia, Ryukyu Islands, Micronesia and Polynesia (Samoa) to subtropical regions of Australia and has been used by the society. This study aimed to determine the proximate compositions, bioactive compounds and antioxidant activity from large-leafed mangrove fruit which extracted by methanol. It have high carbohydrate content is 29.28%, 66.39% moisture, 2.11% protein, 1.07% fat and 1.15% ash. Old fruit yield greater than young fruit with value 9.94% and 6.83%, respectively. Old fruit has a more effective antioxidant activity (13.47 ppm) compared to young fruit (81.60 ppm) and classified as a very strong antioxidant (IC₅₀ < 50 ppm). Bioactive compounds that act as antioxidants are phenol group. By TLC chromatography technique produces the best eluent, namely methanol and water (4:1) and result 3 fractions. Fraction III had the most effective antioxidant activity with IC₅₀ value of 26.69 ppm.

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Introduction

Antioxidants are compounds that can inhibit or prevent the occurrence of oxidation of the substrate is easily oxidized and has been used by society. Limited reserves of antioxidants in the body so that in the event of exposure excess free radicals, the body requires sources of antioxidants that come from outside. Antioxidants are grouped into two categories based on the source, which are natural antioxidant and synthetic antioxidants. Natural antioxidants are antioxidants obtained natural and be a potential alternative to be developed as a replacement synthetic antioxidants (Winarsi, 2007). Natural antioxidants contain bioactive compounds.

Abstract

Bioactive compounds can be determined by phytochemical tests. These compounds act as a stamina strengthen, immune system and prevent several diseases, such as cancer, heart disease, stroke, high blood pressure, cataract, osteoporosis and infections digestive tract. Phytochemical compounds found in plants, namely alkaloids, flavonoids, quinones, tannins, polyphenols, saponins, steroids and triterpenoids (Juniarti *et al.*, 2009). One of the plants as a potential source of bioactive compounds is large-leafed mangrove (*Bruguiera gymnorrhiza*).

Large-leafed mangrove live in mangrove forest ecosystems. Mangrove forest is widespread areas in

the tropics. Indonesian mangrove forest area reached 50% of the total mangrove Asia and nearly 25% of the world's mangrove forests, which is about 3.7 million Ha (Onrizal, 2010). Mangrove forests as one of the wetlands in the tropics with easy access and usability components of high biodiversity and land has made these resources as resources tropical are threatened sustainability (Onrizal, 2005). This is due to the conversion of mangrove forests continue increased so it is necessary to do research on the utilization of mangrove plants, such as the development of functional foods that can serve as antioxidant, antihypertensive and hepatoprotective.

Large-leafed mangrove plants are found in tropical regions of the Pacific from Southeast Asia, Islands Ryukyu, Micronesia and Polynesia (Samoa) to subtropical regions of Australia and has been used by the society. Fruit of this plants have antiviral activity and can against Sarcoma I80 tumor and Lewis lung carcinoma cancerous and high carbohydrate that has the potential to become a new food source. The bark is used to treat burns in the Solomon Islands, diarrhea and malaria in Indonesia and Cambodia (Allen and Duke, 2006). Compounds that play a role as anticancer, anti-diarrhea and malaria has not been studied scientifically. Compound antioxidants thought to play a role in it.

Research on antioxidants and bioactive

compounds of plants and plant water mangrove has been done, e.g. research velvetleaf plants (Jacoeb *et al.*, 2010), water clover (Nurjanah *et al.*, 2012), water spinach (Nurjanah *et al.*, 2014) and *Sonneratia caseolaris* fruit (Santoso *et al.*, 2011), but research on *B. gymnorrhiza* plant still very little. Research of Jacoeb *et al.* (2013) produced the largest yield of crude extract of large-leafed mangrove (*B. gymnorrhiza*) fruit were extracted with methanol, which is 7.85%. The study also result highest antioxidant activity in methanol extracts with IC₅₀9.42 ppm, but compounds that act as antioxidants are not done isolation and separation (purification) so that the study of this needs to be done.

This study was aims to determine the proximate compositions, antioxidant activity and bioactive compounds as antioxidants function as free radical scavengers from large-leafed mangrove (*Bruguiera gymnorrhiza*) fruit extracted by methanol. The study began with the collection and preparation of samples, proximate analysis, extraction, and separation (purification) using thin layer chromatography (TLC) and column chromatography. Antioxidant activity assay performed before and after separation (purification).

Materials and Methods

Raw materials

The materials used in the study is the largeleafed mangrove (*B. gymnorrhiza*) fruit, methanol pa. (extraction). and materials for separation of bioactive compounds and antioxidant activity assay (free radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) Alorich Chem. and Ascorbic acid HmBG Chem.) and phytochemical test. Tools used, i.e. orbital shaker WiseShake, rotary vacuum evaporator Rot Buchi R-205, UV-Vis spectrophotometer Hitachi U-2800, separation by Thin Layer Chromatography (TLC) and Column Chromatography.

Sample preparation and extraction

The study consisted of two stages. The first stage are the collection and preparation of sample, proximate analysis, extraction, phytochemical test and antioxidant activity assay of crude extracts. The second stage are the separation or purification and antioxidant activity test of fractions (results purification). Its fruit used, the old fruits (fruit greenbrown, reddish brown petals) and young (green fruit, green tanned petals). Preparation process includes cleaning and drying fruit. Fruits what had clean to reduced size and dried in the sun for 3 days. Fruits that has been dried to obtain a powder of crushed dried and stored in plastic at chilling temperatures for process next research.

Powder of fruits dried were extracted using methanol with the method maceration at room temperature with orbital shaker at 150 rpm. Comparison of sample and solvent is 1:4 (w/v). Extraction carried out for 48 hours and then performed using paper filtration Whatman 42 (Hardiningtyas, 2012). The extraction process is not done until the filtrate colored (clear). The filtrate is collected and then evaporated using a rotary vacuum evaporator at 40°C and then dried using a freeze dryer thus obtained crude extracts of fruit in solid form.

Proximate analysis

Proximate compositions of large-leafed mangrove determined by proximate analysis. Proximate test performed based on AOAC (2005). Analysis was conducted on the test moisture content, fat, protein and ash. Carbohydrate content obtained by difference.

Antioxidant activity assay

Antioxidant activity determined DPPH method by Hanani *et al.* (2005). DPPH test method is one of the methods most widely used to determine the efficiency of the performance of a substance that acts as a antioxidants (Molyneux, 2004). The early stages of testing antioxidant activity is prepare the test solution. Crude extract of samples were dissolved in methanol with concentrations of 10, 20, 30 and 40 ppm. Ascorbic acid was used as positive control and comparison with the concentration of 2, 4, 6 and 8 ppm.

DPPH solution concentration used was 1 mM. The solution used in fresh condition and protected from light. A total of 4.5 ml of test solution included in a test tube is then reacted with 0.5 ml of DPPH solution. Test tube is covered with aluminum foil and incubated at 37°C for 30 minutes then the absorbance was measured using a UV-Vis spectrophotometer at length wave 517 nm.

The antioxidant activity of each sample was expressed in percentage inhibition of free radicals which is calculated by the formula:

$$\% inhibition = \frac{blanko \ absorbance - Sample \ absorbance}{blanko \ absorbance} \ge 100\%$$

Concentration and barriers to extract the value of each plotted on the x axis and y. Obtained equation in the form [y = b(x) + a] is used to find the value of IC (Inhibitory concentration) with a stated value of y is 50 and the value of x as IC₅₀. IC₅₀ values is the concentration of the sample solution is required to reduce DPPH by 50%. This test is performed

three replications. The data obtained analyzed descriptively.

Phytochemical test

Phytochemical testing conducted to determine the bioactive components contained in the crude extract of the fruit elected. Phytochemicals test based on Harborne (1987) include alkaloids, steroids, flavonoids, phenols hydroquinone and tannin.

Separation of the bioactive compounds

Separation of the bioactive compounds selected fruit extracts done in two stages separation based on Sarker *et al.* (2006). The first stage is the separation by thin layer chromatography (TLC) which aims to determine the appropriate eluent to separate the active compounds in the crude extract. The second stage is the separation by column chromatography to collecting fractions of active compounds using the appropriate eluent later each fraction was tested antioxidant activity.

Results and Discussion

Proximate compositions

Large-leafed mangrove fruit used by people as a food substitute for rice in when a bad season. The fruit is usually treated with boiled, dried and can be stored for a long time. Proximate compositions of large-leafed mangrove fruit are presented in Table 1. It's that used in the study have the proximate composition is not much different from it fruit used by other researchers, namely Jacoeb *et al.* (2013) and Fortuna (2005). Carbohydrate content is higher than that obtained with fruit Avicennia marina fruit (Table 1). Differences in the chemical composition in a material can be influenced by the environment or habitat and species (Megayana *et al.*, 2012).

Crude extract

Fruit powder were extracted using methanol solvent. Solvent methanol selected based on research conducted by Jacoeb *et al.* (2013). The study resulted in the largest yield and antioxidant activity effective on large-leafed mangrove fruit extracted using methanol solvent. Prior *et al.* (2005) which states that methanol is a polar compound that is easy to position the atoms hydrogen from a compound or hydroxyl groups to form hydrogen bonds so as to facilitate movement of protons (hydrogen atoms antioxidants) to free radicals. Extract the results obtained dark brown. Old fruit yield (9.94%) has a value greater than young fruit (6.83%). It can caused by different harvesting. Harvesting is one of the factors that affect the chemical composition or content of bioactive

Table 1. Proximate composit	tion of fruit large-leafed
mangrove (B. gymnorrhiza)) and Avicennia marina

Proximate	Bruguiera gymnorrhiza			Avicennia marina
composition	Current result	Jacoeb et al. (2013)	Fortuna (2005)	Hardiningtyas (2012)
(wet basis)	(%)	(%)	(%)	(%)
Moisture	66.39	62.92	73.76	68.16
Ash	1.15	1.29	0.34	4.45
Crude protein	2.11	2.11	1.13	3.67
Crude lipid	1.07	0.79	1.25	0.72
Carbohydrate	29.28	32.91	23.53	23.00

Table 2. Antioxidant activity of crude extract and ascorbic

acid			
Sample	Linear line	IC50 value (ppm)	Antioxidant1
Crude extract			
Old	y = 0.86 + 38.41	13.46	Very strong
Young	y = 0.22x + 32.13	81.60	Strong
Ascorbic acid	y = 7.01x + 22.77	3.88	Very strong
¹ Molyneu	ix (2004)		

Table 3. TLC result of crude extract by methanol:water

Spots	Solvent (cm)	Compound (cm)	Rf value
I	8.00	2.50	0.31
II	8.00	2.70	0.34
III	8.00	3.60	0.45

Table 4. Antioxidant activity of fraction				
	Sample	Linear line	IC50 value (ppm)	Antioxidant1
	Fraction I	y = 1.48x + 0.58	33.50	very strong
	Fraction II	y = 1.28x + 0.45	38.79	very strong
	Fraction III	y = 2.17x - 7.99	26.69	very strong
	¹ Molyneux (2004)			

materials (Megayana *et al.*, 2012). Yield results that are suspected to be high content of bioactive found in old fruits.

The results obtained are supported by research conducted Khaerana *et al.* (2008) which states that the impact of harvesting on the content xanthorrhizol on ginger rhizome. The content of these compounds generally increased in plants were harvested at 7 months compared to 5 months of age were harvested. This result is also in accordance with research conducted by Megayana *et al.* (2012) which produces content of Na-alginate seaweed *Sargassum* sp. harvest at the age of 2 weeks (5.54%) increasing in the harvest age 6 weeks (10.62%).

Antioxidant activity of crude extract

Antioxidant activity of the test method used in this study is the method DPPH. This method is the most widely used method for estimate the effectiveness of the performance of substance that acts as an antioxidant. This method chosen because it is a method that is simple, easy, fast, sensitive and only require the least amount of material. Antioxidant activity of large-leafed mangrove crude extract and ascorbic acid presented in Table 2. Old fruits has a higher antioxidant activity than young fruit. This results proves that the extract yield contains a bioactive component obtained high. Old fruit and ascorbic acid has a very strong antioxidant activity, while the young fruits including strong antioxidants (Table 2).

Bioactive compounds of crude extract

Phytochemical content of the methanol extract of the large-leafed mangrove fruit is determined by phytochemical test. Phytochemical compounds contained in the crude extract of the large-leafed mangrove fruit are steroids, flavonoids, phenols hydroquinone and tannin.

Phenolic compounds are compounds that are soluble in polar compounds and slightly polar. Phenolic compounds can be flavonoids, phenols simple monocyclic, phenyl propanol and phenolic quinones (Harborne, 1987). Prasad et al. (2009) states that flavonoids are a group of natural compounds of the most diverse and dispersed wide. These compounds have chemical and biological activity spectrum broad including the scavenging of free radicals. Wu et al. (2004) states that the antioxidant a phenolic components work easily in donating a hydrogen atom the peroxyl radical (ROO*). Valgimigli et al. (1995) adds that the effectiveness reduction of free radicals by polyphenols is affected number of hydroxyl groups on the compound. Prior et al. (2005) asserts that the effectiveness and strength of activity antioxidant is determined by the ability to move the hydrogen atom and transfer of a single electron.

Antioxidant compounds separation by chromatography

Chromatographic techniques are used for separation by thin layer chromatography (TLC) and column chromatography. TLC technique used to determine the solution developers (eluent). Obtained the best eluent for the separation of crude extract of large-leafed mangrove fruit parents of methanol:water (4:1). This results in the separation of compounds with 3 fractions point (spotting) are different. TLC results can be seen in Table 3. Eluent is then used in the chromatography column. The separation yield 3 fractions based on the same pattern of spots. Fraction is then freeze-dried (freeze dry) so that the fraction obtained in solid form. Fraction then tested the antioxidant activity.

Antioxidant activity of fractions

Solid fractions obtained from the separation tested antioxidant activity with concentrations of 10, 20, 30 and 40 ppm. The test results presented in Table 4 that produces fraction III as a fraction whose activity best antioxidants, which amounted to 26.69 ppm. The value of antioxidant activity differences between the crude extract and fractions. Difference might be due to the antioxidant activity the effect of synergism between the active compounds contained in fruit extracts so that when the compound is separated, then the activity decreased. This case supported by Moure *et al.* (2001), that the combined antioxidant components produce synergistic antioxidant effectiveness higher than the activity in a single component. Mu *et al.* (2007), also produce a combination components of flavonoids and triterpenoids which have neuroprotective effects, antioxidant activity and anti-inflammatory more effective than the single components. Research Santos *et al.* (2010) produce effects synergism between the components quinonemetida flavonoids and triterpenes in providing antioxidant effects.

Conclusions

Large-leafed mangrove (*B. gymnorrhiza*) fruits have high carbohydrate content that is 29.28%, moisture content 66.39%, 2.11% protein, 1.07% fat and 1.15% ash. Old fruit yield more than young fruits with values 9.94% and 6.83%, respectively. Old fruit has a more effective antioxidant activity (13.47 ppm) than younger fruit (81.60 ppm) and classified as a very strong antioxidant (IC₅₀ <50 ppm). Classes of active compounds that act as antioxidants are phenol group. By TLC chromatography technique produces the best eluent, namely methanol:water (4:1) and yielded three fractions. Best fraction (fraction III) have antioxidant activity is 26.69 ppm.

References

- Allen, J.A. and Duke, N.C. 2006. *Bruguiera gymnorrhiza* (large-leafed mangrove). Download from *http://agroforestry.net/tti/B.gymno-largeleafmangrove.pdf*.
- [AOAC] Association of Official Analytical Chemist. 2005. Official Method of Analysis of of the Association of Official Analytical Chemist 18th Edition. Arlington: The Association of Official Analytical Chemist, Inc.
- Fortuna, J. 2005. Founded fruit mangroves as a staple food. Download from *http://tempo.co.id/hg/nusa/ nusatenggara/* 2005/09/08/brk,20050908-66318,id. *html.*
- Hanani, E., Moneim, B. and Sekarini, R. 2005. Identification of antioxidant compounds in the sponge *Callispongia* sp of the Thousand Islands. Magazine Pharmaceutical Sciences 2 (3): 127-133.
- Harborne, J.B. 1987. Methods Phytochemicals 2nd Edition.
 Padmawinata K, Soediro I, translator. Bandung: Bandung Institute of Technology. Translation of: Phytochemicals Methods.
- Hardiningtyas, S.D. 2012. Antioxidant activity and hepatoprotective effects of fires leaves white (*Avicennia marina*). Bogor, Indonesia: Bogor Agricultural University, M.Sc thesis.
- Jacoeb, A.M., Abdullah, A. and Rushdi, R. 2010. Microscopic characteristics and components bioactive plant velvetleaf (*Limnocharis flava*) from Situ Gede, Bogor. Journal Aquatic 4 (2): 1-6.
- Jacoeb, A.M., Suptijah, P. and Zahidah. 2013. Bioactive components and antioxidant activity of large-leafed mangrove fruit (*Bruguiera gymnorrhiza*). Journals Indonesian Fishery Product Processing 16(1): 86-94.

- Juniarti, Osmeli, D. and Yuhernita. 2009. The content of chemical compounds, toxicity tests (Brine Shrimp Lethality Test) and antioxidants (1,1-diphenyl-2-picrilhydrazyl) of the extract sage leaves (*Abru sprecatorius* L.). Makara Science 13 (1): 50-54.
- Khaerana, Ghulamahdi, M. and Purwakusumah, E. D. 2008. Effect of drought stress and harvesting on growth and content of ginger xanthorrhizol (*Xanthorriza curcuma* Roxb.). Agronomy Bulletin 36 (3): 241-247.
- Megayana, Y., Subekti, S. and Alamsjah, M.A. 2012. Studies and chlorophyll content of alginate seaweed *Sargassum* sp. at different harvest time. Journal of Aquaculture and Fish Health 1 (1): 120-127.
- Molyneux, P. 2004. The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. Journal of Science of Technology 26(2): 211-219.
- Moure, A., Cruz, J.M., Franco, D. and Dominguez, J.M., Sineiro, J., Dominguez, H., Nuñez, M.J. and Parajó, J.C. 2001. Natural antioxidants from residual sources. Food Chemistry 72: 145-171.
- Mu, L., Kou, J., Zhu, D. and Yu, B. 2007. Comparison of neuroprotective effects of flavonoids, terpenoids, and their combinations from Ginkgo biloba on ischemiareperfusion-injured mice. Pharmaceutical Biology 45 (9): 728-733.
- Nurjanah, Azka, A. and Abdullah, A. 2012. Antioxidant activity and active components clover water (*Marsilea crenata*) Innovation and Entrepreneurship Journal 1 (3): 152-158.
- Nurjanah, Abdullah, A., Sudirman, S. 2014. Antioxidant activity and active components water spinach (*Ipomoea aquatica* Forsk.). Journal of Innovation and Entrepreneurship 3 (1): 68-75.
- Onrizal. 2005. Mangrove forests to save the people on the north coast of Nias tsunami. Wetlands Conservation News 13 (2): 5-7.
- Onrizal. 2010. Changes in mangrove forest cover in the East Coast of North Sumatra period from 1977 to 2006. Indonesian Journal of Biology 9 (2): 163-172.
- Prasad, K.N., Yang, B., Dong, X., Jiang, G., Zhang, H., Xie, H. and Jiang, Y. 2009. Flavonoid contents and antioxidant activities from *Cinnamomun* species. Innovative Food Science and Emerging Technologies 10: 627-632.
- Prior, R.L., Wu, X. and Schaich, K. 2005. A standardized method for the determination of antioxidant capacity and phenolic in foods and dietary supplements. Journal of Agricultural and Food Chemistry 53: 4290-4302.
- Santos, V.A.F.F., Santos, D.P., Gamboa, I.C., Zanoni M.V.B. and Furlan, M. 2010. Evaluation of antioxidant capacity and synergistic associations of quinonemethide triterpenes and phenolic substances from *Maytenus ilicifolia* (Celastraceae). Molecules 15: 6956-6973.
- Santoso, J., Febrianti, F. and Nurjanah. 2011. Phenol content, chemical composition and activity antioxidant Sonneratia caseolaris fruit. Journal of Food Science and Technology 9 (1): 1-10.

- Sarker, S.D., Latif, Z. and Gray, A.I. 2006. Natural Product Isolation. Inside: Sarker SD, Latif Z, Gray AI. (Eds).
 Methods in Biotechnology Natural Product Isolation.
 2nd edition. New Jersey: Humana Press Inc.
- Valgimigli, L., Banks, J.T., Lusztyk, A. and Ingold, K.U., 1995. Kinetic solvent effects on hydroxyl hydrogen atom abstractions are independent of the nature of the abstracting radical. Two extreme test using vitamin E and Phenol. Journal of the American Chemical Society 117: 9966-9971.
- Winarsi, H. 2007. Antioxidants and Free Radicals Natural. Yogyakarta, Indonesia: Kanisius.
- Wu, X., Beecher, G.R., Holden, J.M., Haytowitz, D.B., Gebhardt, S.E. and Prior, R.L. 2004. Lipophilic and hydrophilic antioxidant capacities of Common Foods in the United States. Journal of Agriculture and Food Chemistry 52: 4026-4037.